Acta Veterinaria-Beograd 2017, 67 (2), 168-177 UDK: 636.2.09:618.19-002

DOI: 10.1515/acve-2017-0015

Research article

DETECTION OF *icaA* AND *icaD* GENES OF *Staphylococcus* aureus ISOLATED IN CASES OF BOVINE MASTITIS IN THE REPUBLIC OF SERBIA

SUVAJDŽIĆ Branko*, TEODOROVIĆ Vlado, VASILEV Dragan, KARABASIL Neđeljko, DIMITRIJEVIĆ Mirjana, ĐORĐEVIĆ Jasna, KATIĆ Vera

Department of Food Hygiene and Technology of Animal Origin, Faculty of Veterinary Medicine, University of Belgrade, 11000 Belgrade, Serbia

(Received 22 June, Accepted 29 December 2016)

Staphylococcus aureus (S. aureus) is the most common agent of contagious mastitis that causes serious health and economic problems. The ability to form biofilms is an important virulence factor of S. aureus for the establishment of persistent infections. This study is aimed to investigate the presence of icaA and icaD, two genes of importance for the biofilm formation in S. aureus bovine mastitis isolates. In order to isolate and identify S. aureus, 1555 milk samples were collected from 401 cows, located in different regions of the Republic of Serbia. Using the conventional microbiological methods 100 isolates were characterized as coagulase-positive staphylococci. After primary biochemical identification, the species confirmation of 44 S. aureus isolates was done using MALDI-TOF Mass Spectrometry and PCR technique, targeting the S. aureus-specific nuc gene. Among all investigated S. aureus isolates 25.0% harbored both icaA gene and icaD genes. The presence of icaD gene alone was confirmed in 40.9% of cases totaling icaD positive isolates to 65.9%. The remaining 34.1% of S. aureus isolates were negative for the presence of both genes. The results of the present study indicate the existence of potential biofilm-producer strains in different regions of the Republic of Serbia, both under intensive and semi-extensive cows breeding.

Key Words: biofilm, icaA gene, icaD gene, mastitis, Staphylococcus aureus

INTRODUCTION

Mastitis is the most important disease in dairy cows leading to substantial economic losses in the primary milk production worldwide [1-2]. *S. aureus* has been described as a major pathogen responsible for this disease among more than 137 different microorganisms associated with the etiology of mastitis [3,4]. The infected mammary gland is the main reservoir of *S. aureus* while the transmission between cows usually occurs during milking [5]. Early detection and characterization of *S. aureus* strains are very important for mastitis prevention and management [6]. In veterinary microbiology,

^{*}Corresponding author: e-mail: brankos@vet.bg.ac.rs

several techniques are used for the identification and the characterization of *S. aureus* mastitis isolates. The routine microbiological methods applied to microorganism identification require a lot of time and use of various biochemical tests, as well as the evaluation of experts [7]. Compared to the conventional phenotypic methods, molecular methods provide faster and more accurate responses during the identification of the microorganisms [8,9]. MALDI-TOF Mass Spectrometry and PCR are valuable in the rapid identification of this contagious mastitis agent while the PCR technique is also used to test the pathogenic potential of *S. aureus* mastitis isolates.

Many virulence factors are involved in S. aureus confrontation of the mammary gland defense system. The ability to form biofilm is an important virulence factor of S. aureus in the pathogenesis of mastitis [10,11]. Biofilm formation is a very complex process that begins by adhesion of S. aureus to the tissue surface, using surfaceanchored proteins which bind to host matrix proteins (MSCRAMMs), exposed due to tissue damage [12,13]. After primary adhesion, bacterial cells multiply and produce an extracellular biofilm matrix, also known as slime, which provides interactions between bacterial cells. The main component of the S. aureus slime is the exopolysaccharide poly-N-acetyl-β-1,6-glucosamine (PNAG), synthesized by enzymes encoded in the intercellular adhesion (ica) locus [14]. The ica locus belongs to the "accessory genes" of the genome [15] and contains icaA, icaB, icaC and icaD genes, of which icaA and ivaD play a significant role in S. aureus biofilm formation [16]. The ivaA gene encodes enzyme N-acetylglucosaminyl transferase whose enzymic activity becomes significant only in the case of icaD gene expression [15]. The product of icaC gene is involved in the translocation of the poly-N-acetylglucosamine molecule to the surface of the bacterial cell while iaB gene encodes the surface-attached protein which performs the deacetylation reaction essential for the interactions between bacterial cells and promotes biofilm development [15,17].

Detection of the *ivaA* and *ivaD* genes using PCR assay is reliable for determining the potential of *S. aureus* isolates to produce biofilms and may help in the rapid detection of biofilm-producer strains [6]. Therefore, the aim of this study was to investigate the frequency of the biofilm producing related genes, *ivaA* and *ivaD*, occurrence in *S. aureus* mastitis isolates originated from different regions of the Republic of Serbia.

MATERIALS AND METHODS

Milk sampling

A total of 1555 milk samples were collected from 401 cows from different regions of the Republic of Serbia. Milk samples were collected from udder quarters with increased somatic cell counts, as determined by the California mastitis test or from cows with clinical mastitis. Before milking the udder skin was cleaned, washed and dried. A volume of 10 ml of milk per udder quarter was collected in sterile properly labeled tubes, after disinfection of teat ends with 70% ethanol. Milk samples were kept

at 4°C and transported to the laboratory of the Department for Food Hygiene and Technology of Animal Origin, Faculty of Veterinary Medicine, University of Belgrade.

Microbiological assessment

The isolation of *S. aureus* was carried by streaking 0,1 mL of each milk sample, respectively, onto the Columbia agar plates supplemented with 5% sheep blood followed by 24h incubation at 37°C under aerobic conditions. After incubation, typical colonies were subjected to Gram-staining and positive isolates subcultured on Trypticase soy agar 24h at 37°C for catalase and coagulase testing. API Staph-Ident system (BioMérieux, France) was used for biochemical identification of coagulase-positive staphylococci. 44 selected *S. aureus* isolates were then subjected to the species confirmation using *MALDI-TOF Mass Spectrometry* and *PCR*.

MALDI-TOF Mass Spectrometry

The species confirmation of 44 isolates was done using Vitek MS (bioMérieux, France) based on MALDI-TOF Mass Spectrometry technology. Preparation of investigated isolates was done according to the manufacturer. Escherichia coli ATCC® 8739 was used for calibration of the apparatus while VITEK MS V2.0 Knowledge Base – Industry Use was used for results reading.

DNA extraction

Extraction of genomic DNA was carried out by the boiling method. *S. aureus* isolates were subcultured on Trypticase soy agar at 37°C for 24h. After the incubation period, a fresh colony of each isolate was suspended in 500 µL of DNase-RNase free water. The suspension was held in a water bath at 100°C for 10 min and then cooled on over the next 10 min. After centrifugation at 10 000 rpm for 5 min, the supernatant containing bacterial DNA was used as a template for PCR amplification.

PCR assays

The molecular identification (species confirmation) of 44 chosen isolates was done using PCR targeting S. *aureus*-specific *nuc* gene (encoding thermostable nuclease). Primers used for the amplification of the *nuc* gene are shown in Table 1. All PCR reactions were performed in 50 μL reaction volumes containing 25 μL Dream Taq Master Mix (2X) that contains 2× Dream Taq buffer, 4 mM MgCl₂ and 0,4 mM of each of the 4 dNTPs (Thermo Scientific, Lithuania), 1 μL of each primer, 1μL of template DNA and nuclease free water to 50 μL. DNA amplification was performed in a FlexCycler (AnalyticJena, Germany). The program for amplification of *nuc* gene was: (1) initial denaturing step at 95°C for 5 min; (2) 30 cycles of 30 s at 95°C, 30 s at 55°C, and 1 min at 72°C; and (3) final extension step at 72°C for 7 min. Amplification products were analyzed by electrophoresis on 1.5% (wt/vol) agarose (TopVision

agarose Thermo Scientific, Lithuania) gel, after staining with ethidium bromide. *S. aureus* ATCC 25923 was used as a positive control while Gene Ruler 100 bp DNA ladder (Thermo Fisher Scientific, Lithuania) was used as the molecular weight marker.

In order to determine the potential of *S. aureus* isolates to produce slime, PCR assay targeting *ivaA* and *ivaD* genes was performed [13]. Primers used for the amplification of *ivaA* and *ivaD* genes are shown in Table 1. All primers used in this study were obtained from Invitrogen (United States). Ten microliters of the extracted DNA was used as a template in a 50 μL PCR mixture, that contained 25 μL Dream Taq Master Mix (2X) (Thermo Scientific, Lithuania), 5 μL of each primer, and nuclease free water to 50 μL. The amplification of *ivaA* and *ivaD* gene was performed as follows: (1) initial denaturing step at 92°C for 5 min; (2) 30 cycles of 92°C for 1 min, 49°C for 50 s and 72°C for 1 min; and (3) final extension step at 72°C for 7 min. *S. aureus* ATCC 6538 was used as a positive control for both genes [21] and Gene Ruler 100 bp DNA ladder as the molecular weight marker. The PCR for *ivaA* and *ivaD* genes have amplified the products of 1315 bp and 381 bp, respectively (Figure 1). Electrophoresis was performed in 1.5% (wt/vol) agarose gel stained with ethidium bromide.

Gen	Primer	Sequence (5'-3')	Reference
Nuc	nuc - f	TCA GCA AAT GCA TCA CAA ACA G	[8]
	nuc - r	CGT AAA TGC ACT TGC TTC AGG	
IcaA	icaA - f	CCT AAC TAA CGA AAG GTA G	[13, 17]
	icaA - r	AAG ATA TAG CGA TAA GTG C	
IcaD	icaD - f	AAA CGT AAG AGA GGT GG	[13, 17]
	icaD - r	GGC AAT ATG ATC AAG ATA C	

Table 1. Primers used for the amplification of *nuc*, *icaA* and *icaD* genes

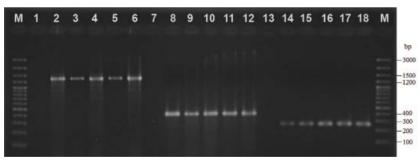


Figure 1. Agarose gel electrophoresis with PCR amplicons of *icaA*, *icaD* and *nuc* gene Legend: M: molecular weight marker; 1: negative control for *icaA* gene; 2-5: *S. aureus icaA* (1315 bp) positive isolates; 6: positive control for *icaA* gene (*S. aureus* ATCC 6538); 7: negative control for *icaD* gene; 8-11 *S. aureus icaD* (381 bp) positive isolates; 12: positive control for *icaD* gene (*S. aureus* ATCC 6538); 13: negative control for *nuc* gene; 14-17: *S. aureus* isolates - species confirmation based on *nuc* gene (255 bp); 18: positive control for *nuc* gene (*S. aureus* ATCC 25923).

RESULTS

After conventional microbiological testing, 100 isolates of coagulase-positive staphylococci were isolated from 1555 milk samples. Further biochemical and molecular identification confirmed 44 selected isolates as *S. aureus* with the presence of species specific *nuc* gene in all examined isolates (Figure 1 and data not shown).

Among all investigated *S. aureus* isolates, 11 out of 44 (25.0%) harbored both *icaA* and *icaD* genes. The presence of *icaD* gene alone was observed in 18 out of 44 (40.9%) *S. aureus*, isolates, concluding the total number of isolates with *icaD* gene to 29 (65.9%). The remaining 15 (34.1%) *S. aureus* isolates did not harbor any of the tested genes (Figure 2).

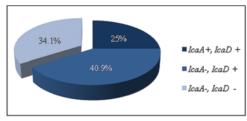


Figure 2. Results of icaA and icaD genes detection in S. aureus mastitis isolates

DISCUSSION

Due to the various virulence factors, S. aureus successfully opposes the mammary gland defense system leading to the development of the pathological process. The ability of S. aureus to form biofilm is an important factor in the mastitis pathogenesis that enables the long-term persistence of bacteria in the mammary glands, leading to the chronic form of the disease [10,19]. This ability is often associated with the decreased efficacy of antibiotic therapy and problems with infection eradication [20]. Rapid detection of biofilm producing S. aureus strains along with the knowledge of biofilm formation mechanisms enables early application of the proper corrective measures [21]. The most studied biofilm mechanism in S. aureus is ica operon dependent mechanism and previous research demonstrated that the majority of clinical isolates, both from human and bovine mastitis contain the ica operon [22]. Therefore, PCR assay standardized for detection of the icaA and icaD genes can determine the potential of S. aureus isolates to produce biofilm and may help in the rapid detection of biofilm-producer strains [6]. Among 44 confirmed S. aureus isolates investigated in this study, only 11 (25.4%) were positive for both icaA and icaD gene. Similar results were reported by Ciftci et al. [23] who determined the presence of both ita A and ita D gene in 25% of S. aureus mastitis isolates while Dhanawade et al. [24] have found 35.29% of S. aureus isolates positive for the presence of both genes. In contrast to our findings, Vasudevan et al. [10] and Fabres-Klein et al. [25] have found that all investigated mastitis isolates of S. aureus were positive for both genes. However, 18 of 44 (40.9%) isolates in the present study

were positive only for icaD gene. Arciola et al. [26] were not detected a gene deletion within the ica locus and pointed out that the genes of the ica locus are strictly linked to each other, so they are either all present or all absent. The findings of Ciftci et al. [23] speak in favor of results from this study, giving that they also determined only the presence of icaD gene in a portion of S. aureus mastitis isolates. The absence of icaA gene detected in icaD gene positive S. aureus isolates, the same authors explained with the possibility of mutation in the iva A gene. Using the sequencing analysis, Murugan et al. [27] have found a high similarity, but some differences in the icaA genes sequences of the 24 S. aureus isolates from conjunctivitis patients. Szweda et al. [28] pointed out that some mutations in *ica* genes are possible and this polymorphism may be the reason for the failed amplification of these genes. Considering that used positive control for ivaA gene and 25% investigated isolates were positive during our investigation, we are of the same opinion that the mutations of iva A gene might be the reason for the negative results in our study. Therefore, we speculate that the presence of the icaD gene alone might indicate the existence of the ital locus in a total of 65.9% investigated S. aureus mastitis isolates. Giving that investigated isolates originated from different localities of the Republic of Serbia, the importance of these results is greater.

Distribution of the investigated *S. aureus* isolates is shown in Figure 3. Biofilm-related genes (both *icaA* and *icaD* genes, or *icaD* gene alone) were detected in *S. aureus* isolates originated from 7 out of 10 examined localities. Any potential biofilm-producer strains were not detected in Brasina, Mionica and Paraćin. Both *icaA* and *icaD* genes were not

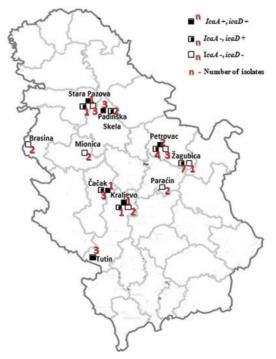


Figure 3. Distribution of the investigated *S. aureus* mastitis isolates

found in all *S. aureus* isolates originated from the same herd (farms located in Stara Pazova, Padinska Skela and Petrovac). This fact indicates the existence of different *S. aureus* strains in the same herd. The tested genes (both *icaA* and *icaD* genes, or *icaD* gene alone) were also detected in *S. aureus* isolates originated from rural households in the municipalities of Čačak, Kraljevo, Tutin and Žagubica. Therefore, results of the present study indicate the existence of potential biofilm-producer strains, both under intensive and semi-extensive cows breeding.

Acknowledgements

The research was supported by the Ministry of Education, Science and Technology Development of the Republic of Serbia, Grant number 31086.

Authors' contributions

BS and VK defined the research theme, gave conception of the research and prepared manuscript. BS and JD did sample collections and laboratory studies. BS carried out the molecular studies. VK, VT, VD, NK and MD participated in the design of the research as well as in the analysis and interpretation of results. All authors read and approved the final manuscript.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

REFERENCES

- 1. Kossaibati MA and Esslemont RJ: The costs of production diseases in dairy herds in England. Vet J 1997, 154:41-51.
- 2. Cantekin Z, Ergun Y, Solmaz H, Özmen GÖ, Demir M, Saidi R: PCR assay with host specific internal control for Staphylococcus aureus from bovine milk samples. Mac Vet Rev 2015, 38:97-100.
- 3. Watts JL: Etiological agents of bovine mastitis. Vet Microbiol 1988, 16:41-66.
- 4. Cojkic A, Cobanovic N, Suvajdzic B, Savic M, Petrujkic B, Dimitrijevic V: Chronic mastitis in cows caused by *Streptococcus dysgalactiae* case report. Vet. Glasnik 2015, 69:301-310.
- 5. Akineden O, Annemuller C, Hassan AA, Lammler C, Wolter W, Zschock M: Toxin genes and other characteristics of *Staphylococcus aureus* isolates from milk of cows with mastitis. Clin Diagn Lab Immunol 2001, 8:959–964.

- Melo PC, Ferreira LM, Filho AN, Zafalon, Vicente HIG, de Souza V: Comparison of methods for the detection of biofilm formation by *Staphylococcus aureus* isolated from bovine subclinical mastitis. Braz J Microbiol 2013, 44:119-124.
- 7. NMC. 1999. Current Concepts of Bovine Mastitis. 4th ed. NMC Inc., Madison, WI.
- 8. Kolbert CP, Persing DH: Ribosomal DNA sequencing as a tool for identification of bacterial pathogens. Curr Opin Microbiol 1999, 2:299–305.
- Barreiro JR, Ferreira CR, Sanvido GB, Kostrzewa M, Maier T, Wegemann B, Böttcher V, Eberlin MN, dos Santos MV: Short communication: Identification of subclinical cow mastitis pathogens in milk by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. J Dairy Sci 2010, 93:5661-5667.
- 10. Vasudevan P, Nair MKM, Annamalai T, Venkitanarayanan KS: Phenotypic and genotypic characterization of bovine mastitis isolates of *Staphylococcus aureus* for biofilm formation. Vet Microbiol 2003, 92:179-185.
- 11. Castelani L, Pilon LE, Martins T, Pozzi CR, Arcaro JRP: Investigation of biofilm production and *icaA* and *icaD* genes in *Staphylococcus aureus* isolated from heifers and cows with mastitis. Anim Sci J 2015, 86:340–344.
- 12. Cucarella C, Solano C, Valle J, Amorena B, Lasa I, Penade JR: Bap, a *Staphylococcus aureus* surface protein involved in biofilm formation. J Bacteriol 2001, 183:2888–2896.
- 13. Foster TJ, Geoghegan JA, Ganesh VK, Höök M: Adhesion, invasion and evasion: the many functions of the surface proteins of *Staphylococcus aureus*. Nat Rev Microbiol 2014, 12:49-62.
- 14. Cramton SE, Gerke C, Schnell NF, Nichols WW, Götz F: The intercellular adhesion (ica) locus is present in *Staphylococcus aureus* and is required for biofilm formation. Infect Immun 1999, 67:5427-5433.
- 15. Arciola CR, Campoccia D, Ravaioli S, Montanaro L: Polysaccharide intercellular adhesin in biofilm: structural and regulatory aspect. Front Cell Infect Microbiol 2015, 5:1-10.
- Arciola CR, Baldassarri L, Montanaro L: Presence of ica A and icaD genes and slime production in a collection of staphylococcal strains from catheter-associated infections. J Clin Microbiol 2001, 39:2151-2156.
- 17. Vuong C, Voyich JM, Fischer ER, Braughton KR, Whitney AR, DeLeo FR., et al.: Polysaccharide intercellular adhesin (PIA) protects *Staphylococcus epidermidis* against major components of the human innate immune system. Cell Microbiol 2004, 6:269–275.
- 18. Ferreira AA, Tette PAS, Mendonça RCS, Soares ADS, Carvalho MMD: Detection of exopolysaccharide production and biofilm-related genes in *Staphylococcus* spp. isolated from a poultry processing plant. Food Sci Technol 2014, 34:710-716.
- 19. Fox LK, Zadoks RN, Gaskins CT: Biofilm production by *Staphylococcus aureus* associated with intramammary infection. Vet Microbiol 2005, 107:295–299.
- 20. Melchior MB, Fink-Gremmels J, Gaastra W: Comparative assessment of the antimicrobial susceptibility of *Staphylococcus aureus* isolates from bovine mastitis in biofilm versus planktonic culture. J Vet Med B Infect Dis Vet Public Health 2006, 53:326-332.
- 21. Oliveira M, Bexiga R, Nunes SF, Carneiro C, Cavaco LM, Bernardo F, et al.: Biofilm-forming ability profiling of *Staphylococcus aureus* and *Staphylococcus epidermidis* mastitis isolates. Vet Microbiol 2006, 118:133–140.
- 22. O'Gara JP: Ica and beyond: biofilm mechanisms and regulation in Staphylococcus epidermidis and *Staphylococcus aureus*. FEMS Microbiol Lett 2007, 270:179–188.

- Ciftci A, Findik A, Onuk EE, Savasan S: Detection of methicillin resistance and slime factor production of *Staphylococcus aureus* in bovine mastitis. Braz J Microbiol 2009, 40:254-61.
- 24. Dhanawade, NB, Kalorey DR, Srinivasan R, Barbuddhe SB, Kurkure NV: Detection of intercellular adhesion genes and biofilm production in *Staphylococcus aureus* isolated from bovine subclinical mastitis. Vet Res Commun 2010, 34:81-89.
- Fabres-Klein MH, Santos MJC, Klein RC, de Souza GN, Ribon ADOB: An association between milk and slime increases biofilm production by bovine *Staphylococcus aureus*. BMC Vet Res 2015, 11:3 DOI 10.1186/s12917-015-0319-7.
- 26. Arciola CR, Gamberini S, Campoccia D, Visai L, Speziale P, Baldassarri L, Montanaro L: A multiplex PCR method for the detection of all five individual genes of *ica* locus in *Staphylococcus epidermidis*. A survey on 400 clinical isolates from prosthesis-associated infections: J Biomed Material Res 2005, 75:408-413.
- Murugan K, Usha M, Malathi P, Al-Sohaibani AS, Chandrasekaran M: Biofilm forming multi drug resistant *Staphylococcus* spp. among patients with conjunctivitis. Pol J Microbiol 2010, 59:233-239.
- 28. Szweda P, Schielmann M, Milewski S, Frankowska A, Jakubczak A: Biofilm production and presence of *ica* and *bap* genes in *Staphylococcus aureus* strains isolated from cows with mastitis in the eastern Poland. Pol J Microbiol 2012, 61:65–69.

DETEKCIJA icaA I icaD GENA KOD IZOLATA Staphylococcus aureus IZOLOVANIH U SLUČAJU MASTITISA KRAVA U REPUBLICI SRBIJI

SUVAJDŽIĆ Branko, TEODOROVIĆ Vlado, VASILEV Dragan, KARABASIL Neđeljko, DIMITRIJEVIĆ Mirjana, ĐORĐEVIĆ Jasna, KATIĆ Vera

Kao uzročnik kontagioznih mastitisa, *Staphylococcus aureus* (*S. aureus*) izaziva ozbiljne zdravstvene i ekonomske probleme. Sposobnost *S. aureus* da stvara biofilm se ističe kao važan faktor virulencije za uspostavljanje perzistentnih infekcija mlečne žlezde. Stoga je istraživanje imalo za cilj da ispita prisustvo *icaA* i *icaD* gena kod izolata *S. aureus* izolovanih u slučaju mastitisa krava, kao gena od značaja za formiranje biofilma. U cilju izolacije i identifikacije *S. aureus*, prikupljeno je ukupno 1555 uzoraka mleka poreklom od 401 krave iz različitih regiona Republike Srbije. Primenom konvencionalnih mikrobioloških metoda, 100 izolata je okarakterisano kao koagulaza pozitivne stafilokoke. Nakon primarne biohemijske identifikacije, izvršena je potvrda vrste 44 izolata *S. aureus* primenom MALDI-TOF masene spektrometrije, kao i PCR tehnike utvrđivanjem prisutva *nuc* gena koji je specifičan za vrstu *S. aureus*. Od svih ispitanih izolata *S. aureus*, 25,0% je posedovalo *icaA* i *icaD* gen. Prisustvo samo *icaD* gena je utvrđeno kod 40,9% izolata, tako da je ukupno 65,9% izolata bilo pozitivno na *icaD* gen. Preostalih 34,1% izolata *S. aureus* nije posedovalo ni *icaA*, ni *icaD* gen. Rezultati

ove studije ukazuju na prisustvo potencijalnih biofilm produkujućih sojeva *S. aureus* u različitim regionima Republike Srbije, kako u intenzivnom tako i u poluekstenzivnom uzgoju krava.