

ABERRANT KARYOTYPE OF AN EXPERIMENTAL RABBIT (*ORYCTOLAGUS CUNICULUS*, LINNAEUS, 1758) WITH A HETEROZYGOUS FIRST AUTOSOMAL PAIR

Z. STANIMIROVIĆ\*, B. SOLDATOVIĆ\*, SANDA DIMITRIJEVIĆ\*\*  
MARIJANA VUČINIĆ\*\*\* and D. ĐOKIĆ\*

\*Department of Biology, and \*\*Department of Parasitic Diseases,  
Faculty of Veterinary Sciences, University of Belgrade, Bul. JNA 18,  
11000 Belgrade, Yugoslavia

\*\*\*Institute for Animal Production, Faculty of Agriculture,  
Nemanjina 6, 11080 Zemun, Yugoslavia

(Received, 20 October 1992)

*Histological and cytogenetic analysis was applied to a male from a breeding stock of experimental rabbits (*Oryctolagus cuniculus*). The rabbit originated from the breeding farm of experimental animals at the Faculty of Veterinary Sciences in Ljubljana. It was 6 months old, and weighed 2.5 to 3 kg. Reduced fertility was evident from the small number of mated females which, conceived frequent premature termination of gravidity, and reduced vitality of offspring.*

*The karyotype analysis showed an unusually large chromosome in the first pair of autosomes. It was assumed to arise as a consequence of the amplification of some chromosomal segments, possibly due to an increase in the concentration of food additives in the food consumed.*

*Reduced fertility was interpreted to be the consequence of impaired spermatogenesis, irregular chromosomal segregation during meiosis, and the production of imbalanced gametes.*

**Key words:** karyotype, cytogenetic, chromosome, gamet

#### INTRODUCTION

Ever increasing pollution of the environment imposes the requirement for permanent monitoring, both in controlled and uncontrolled settings, aimed at timely detection of structural and numerical chromosomal aberrations that may potentially impair domestic animal reproduction.

The examination of animals from the breeding stock of a farm of experimental animals originating from the Faculty of Veterinary Sciences in Ljubljana revealed abnormalities in reproduction of a male laboratory rabbit with avital offspring and reduced conception in mated females.

The normal karyotype of the experimental rabbit contains  $2n = 44$  chromosomes and NF values of 80. Out of the diploid number of chromosomes, 34 autosomes are metacentric or submetacentric, 8 of them are acrocentric or telocentric, while the X and Y chromosomes are submetacentric (Melander 1956, Sarkar et al., 1962; Ray and Williams 1966).

A review of the literature shows the appearance of aberrations in the chromosomal complement of mammals (Dave et al., 1965; Ray and Williams 1966; Grop et al., 1971; Matthey 1972, 1973; White 1973 a, b.; Orlov 1974; Capanna et al., 1973, 1977; Capanna 1981; Gropp and Winking 1981; Gropp et al., 1982; Brooker 1982; Said 1986; Tichy and Vučak 1987; Winking et al., 1980; Stanimirović 1992).

These aberrations are primarily of the Robertson type, predominantly in Rodentia. References that would indicate the existence of chromosomal aberrations such as inversion, deletion, etc., are scarce.

In this context, especially interesting are the findings of Bostock et al., (1979), Traut et al., (1984), Agulnik et al., (1988 and 1990), and Stanimirović (1992). They all demonstrate alterations in chromosome structure caused by insertions.

#### MATERIAL AND METHODS

The investigation included rabbits originating from a farm of experimental animals at the Faculty of Veterinary Sciences in Ljubljana. They were 6 months old and weighed from 2.5 to 3 kg.

Among the examined rabbits a male specimen was observed that, yielded a significantly lower number of offspring than other male rabbits the result as reduced conception in mated females and avitability of embryos of offspring. This was the reason for histological and cytogenetic analysis as an approach to clarify the basis for the observed abnormalities.

Material for the cytogenetic analysis was obtained according to the procedure of Hsu and Patton (1969). Chromosomes were stained in 5% Giemsa in phosphate buffered saline, at pH = 6.8, for 5 to 10 minutes.

#### RESULTS AND DISCUSSION

Histologic analysis demonstrated that the testicles were permeated with excessive connective and fat tissue with impaired spermatogenesis.

Cytogenetic analysis revealed a significant number of cells with abnormal structure, notably in the chromatid domain, rarely with isochromatid breakage. Chromatid breaks were always on different chromosomes at different sites. They were particularly frequent in larger autosomes and the X chromosome.

In 97.01% of the cytogenetically analyzed cells we detected the chromosomal set of  $2n = 44$ , where the first set of autosomes exhibited heterozygosity in terms of chromosome size (Figure 1). One of the chromosomes from the first pair of autosomes was unusual dimensions (Figure 2). Based on literature data, we propose that the observed phenome-

non may be the consequence of DNA amplification caused by food additives in the diet consumed. This is in agreement with the findings of Bostock et al., (1979), Aglunik et al., (1988, 1990), and Stanimirović (1992).

Out of the total of 150 cells examined we detected 2 to 3 chromatid breaks in 12% of cases and isochromatid breaks in 4% of cases.



Figure 1. Aberrant karyotype of *Oryctolagus cuniculus*

Bostock et al., (1979) examined the influence of pesticides and some food additives on cultured cells and concluded that these chemical agents induce amplification of chromosome segments and even the appearance of out-of-chromosome structures.

Aglunik et al., (1990) examined a specimen of naturally occurring representatives of the species *Mus musculus*. They discovered one unusually large chromosome from the first pair of autosomes with two insertions, one in the 1C5 and one in the 1E region. These mice, also, had reduced reproductive capability as documented by increased mortality of offspring and reduced conception in mated females.

Reduced fertility of the examined animal was notable through frequent termination of gravidity and reduced viability of the offspring which can be interpreted as the consequences of irregular chromosomal segregation causing generation of imbalanced gamates. This is in agreement with the findings of Groppe et al., (1982); Adolph (1984); Nash et al., (1983); Ruvishij et al., (1986 a, b); Aglunik et al., (1990). The cited authors state that heterozygous individuals carry not only insertions but also other types of structural aberration and that in 85% of cases transmit their aberrant chromosome to their offspring.

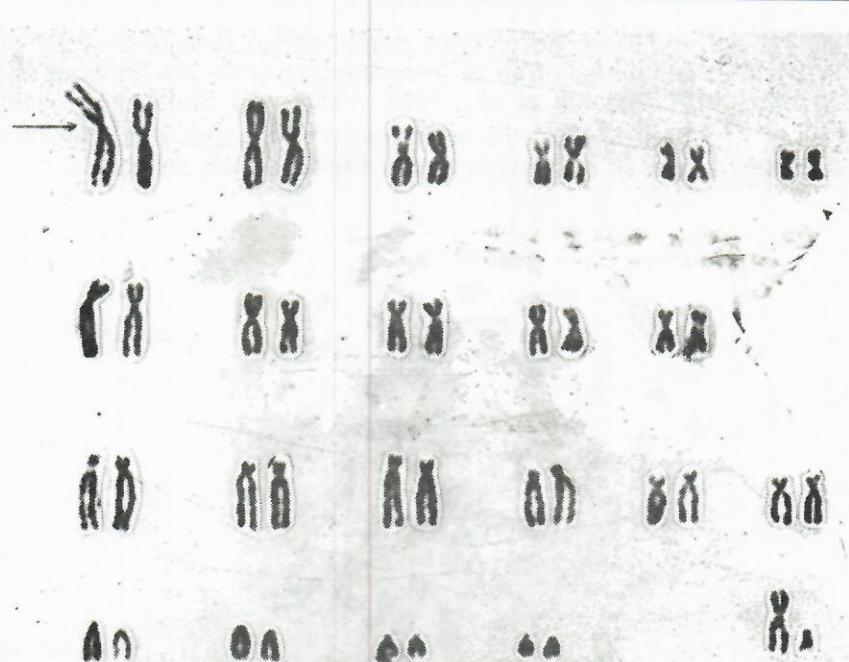


Figure 2. Aberrant karyogram of *Oryctolagus cuniculus* with heterozygous first autosomal pair

#### CONCLUSION

We examined the causes of reduced fertility of a male from a breeding stock of experimental rabbits (*Oryctolagus cuniculus*).

Cytogenetic analysis of the karyotype of the experimental individual indicated heterozygosity in the first pair of autosomes that was reflected in the presence of an unusually large chromosome. Based on literature data we assumed that such a karyotype structure is one of the possible determinants of the impaired fertility that was clinically observed.

The reduced fertility of the examined individual was interpreted to be a consequence of impaired spermatogenesis, irregular chromosomal segregation of the first pair of autosomes, and the production of imbalanced gametes.

#### LITERATURA

1. Adolph von S., 1984. Robertsonsche Translokationen bei Wurtenmbrgischen Hausmausen (*Mus musculus domesticus*) Ein Beispiel zur Chromosomenevolution, *Jh. Ges. Naturkde. Wurttemberg* 139, 67—92.
2. Agulnik S. L., Agulnik A. I., Ruvinskij A. O., 1990. Mejotičeskij dravj aberrentnoj 1-j hromosomi u domovoj miši. *Genetika, Tom 6*, 4, 664—669.
3. Agulnik S. L., Gorlov I. P., Agulnik A. L., 1988. Novij variant 1-j hromozomi u domovoj miši, *Citologija, vol. 30*, 6, 773—778.

4. Bostock C. J., Prescott D. M. and Hatch F. T., 1979. Timing of replication of the satellite DNA-s in cells of kangaroo rat (*Dipodomys aridii*). *Exp. Cell Res.*, 74, 487—495.
5. Brooker P. C., 1982. Robertsonian translocations in *Mus musculus* from N. E. Scotland and Orkney. *Heredity*, 48, 305—309.
6. Capanna E., Civitelli M. V., Cristaldi M., 1973. Chromosomal polymorphism in an Alpine population of *Mus musculus* L., *Boll. Zool.*, 40, 379—383.
7. Capanna E., Civotelli M. V., Cristaldi M. 1977. Chromosomal rearrangement, reproductive isolation and speciation in mammals. The case of *Mus musculus*, *Boll. Zool.*, 44, 213—246.
8. Capanna E., 1981. Caryotype et morphologie crânienne de *Talpa romana* Thomas de terra typica., *Mammalia*, 45, 71—82.
9. Dave, M. J., Takagi, N., Oishi, H. and Kikuchi, Y. 1965. Chromosome studies on the hare and the rabbit. *Proc. Jap. Acad.*, 41—244.
10. Gropp A., Winking H., Muller H. J., Muller J. P., 1971. Chromosomes of the black rat (*R. rattus*), *Mamm. Chromo. News*l., 12, 118—119.
11. Gropp A., and Winking H., 1981. Robertsonian translocations: cytology, meiosis, segregation patterns and biological consequences of heterozygosity, *Symp. zool. Soc. Lond.*, 47, 141—181.
12. Gropp A., Winking H., Redi C., Capanna E., Britton-Davidian J., Noack G., 1982. Robertsonian karyotype variation in wild mice from Rhaeto-Lombardia, *Cytogenet. Cell Genet.*, 34, 67—77.
13. Hsu T. C., Patton J. L., 1969. Bone marrow preparations for chromosome studies. In: Benirschke K.(Ed.) *Comparative mammalian cytogenetics*, Springer Verlag, Berlin, Heiderberg, New York, 454—460.
14. Matthey R. and Jotterand M., 1971. African pygmy mice—the “Robertsonian fan” in the superspecies *Mus minutoides/musculoides*. Karyotypes of two new species, *Mus oubangui*, Petter, *Mus goundae*, Petter., *Mammalian Chromosomes*.
15. Matthey R., 1972. Chromosomes and evolution. *Triangel*, 11, 107—112.
16. Matthey R., 1973. The chromosome formula of eutherian mammals, *Academic Press, London and New York*, 531—616.
17. Melander, Y. 1956. The chromosome complement of the rabbit. *Hereditas* 42, 432.
18. Nach H. R., Brooker P. S., Davis S. J. M., 1983. The Robertsonian translocation house mouse populations of north east Scotland: study of their origin and evolution. *Heredity*, 50, 303—310.
19. Ray, M. and Williams, T. W. 1966. Karyotype of rabbit chromosomes from leucocyte cultures. *Canad. J. Genet & Cytol* 8, 393.
20. Ruvinskij A. O., Agulnik S. I., Agulnik A. I., Beljajev D. K., 1986. Segregacija hromosoma u mišej heterozigotnih po Robertsonovskim translokacijom (Soobćenje 1), *Genetika*, Tom 22, 9, 2326—2332.
21. Ruvinskij A. O., Agulnik S. I., Agulnik A. I., Beljajev D. K., 1986. Segregacija hromosoma u mišej heterozigotnih po Robertsonovskim translokacijam (Soobćenje 2), *Genetika*, Tom 22, 10, 2505—2511.
22. Said K., Jacquot T., Montgelard C., Sonjaya H., Helal A. N., Britton-Davidian J., 1986. Robertsonian house mouse populations in Tunisia: a karyological and biochemical study. *Genetica*, vol. 68, 151—156.
23. Sarker, P., Basu, P. K. and Miller, I. 1962. Karyologic studies on cells from rabbit cornea and other tissues grown in vitro. *Invest. Ophtal.* 1, 33.
24. Sharma, G. P., Parshd, R. and Ghuman, S. K. 1963. On the meiotic and somatic chromosomes of the rabbit. *Oryctolagus cuniculus*, *Res. Bull. (N. S.) Panjab Univ.* 14, 171.
25. Stanimirović Z. 1992. Hromozomski polimorfizam prirodnih populacija *Mus musculus*, Binne (1758) na teritoriji Jugoslavije. *Magistarska teza, Beograd*.
26. Tichy H. and Vučak I., 1987. Chromosomal polymorphism in the house mouse (*Mus domesticus*) of Greece and Yugoslavia, *Chromosoma (Bern)*, vol. 95, 31—36.

27. Traut W., Winking H., Adolph S., 1984. An extra segment in chromosome 1 of wild *Mus musculus*; a C-band positive homogeneously staining region. *Cytogenetics, Cell, Genetics*, vol. 38, 295—297.
28. Winking H., Groop A., Bulfield G., 1981. Robertsonian chromosomes in mice from North-Eastern Greece. *Mouse News Letter*, vol. 64, 69—70.
29. Winking H., Đulić B., Bulfield G., 1988. Robertsonian karyotype variation in the European house mouse, *Mus musculus*, Z., Sugeterkunge, Verlag Paul Parey. Hamburg und Berlin, ISSN, 00443468, 53, 48—161.

**ABERANTNI KARIOTIP LABORATORIJSKOG KUNIĆA (ORYCTOLAGUS CUNICULUS, LINNAEUS, 1758) SA HETEROZIGOTNIM PRVIM PAROM AUTOZOMA**

Z. STANIMIROVIĆ, B. SOLDATOVIĆ, SANDA DIMITRIJEVIĆ,  
MARIJANA VUČINIĆ i D. ĐOKIĆ

**SADRŽAJ**

Histoločkim i citogenetičkim ispitivanjima bio je podvrgnut priplodni mužjak laboratorijskog kunića, (*Oryctolagus cuniculus*) poreklom sa farme eksperimentalnih životinja Veterinarskog fakulteta u Ljubljani starosti oko 6 meseci i telesne mase 2,5—3 kg, zbog smanjene plodnosti, koja se ogledala u smanjenoj konцепцији parenih ženki, učestalom prevremenom prekidu gravida i smanjenoj vitalnosti potomstva.

Analizom kariotipa uočeno je prisustvo jednog neoubičajenog velikog hromozoma iz prvog para autozoma, za koji smo predpostavili da je nastao kao posledica amplifikacija pojedinih delova hromozoma, zbog verovatno povećane koncentracije prehrambenih aditiva u konzumnoj hrani.

Smanjenu plodnost smo tumačili kao posledicu narušene spermatogeneze, neregularne segregacije hromozoma tokom mejoze i produkcije nebalansiranih gameta.