

**The Balkans Scientific Center of the
Russian Academy of Natural Sciences**



1st International Symposium:

**Modern Trends in Agricultural
Production and Environmental
Protection**

PROCEEDINGS

**Tivat-Montenegro
July 02-05.
2019.**

**The Balkans Scientific Center of the
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FUNCTION AND IMPORTANCE OF HSP 70 IN METABOLIC STRESS IN DAIRY COWS IN PERIPARTAL PERIOD

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ABSTRAKT

Peripartal period in dairy cows includes 3 weeks ante partum and 3 weeks post partum (transition period). The transition period represents the most challenging phase in the life of dairy cows, because the body occurring metabolic, endocrine, immune and reproductive changes that affect the animal health and production efficiency. During the transition period (the late gestation and early lactation) in response to metabolic stress, in dairy cows can develop numerous pathophysiological mechanisms (inflammation, insulin resistance and metabolic adaptation). Numerous mechanisms are activated in the organism of dairy cows, the main role of which is maintenance of homeostatic and homeostatic functions of all tissues, organs and organ systems. In regulation of these processes, heat stress proteins HSP70 play a key role.

Key words: *dairy cows, peripartal period, metabolic stress, HSP 70.*

INTRODUCTION

Peripartal period includes 3 weeks ante partum and 3 weeks post partum (the so-called transition period). It represents a period in which the metabolism of dairy cows faces a series of changes in the homeostasis with endocrine changes, metabolic stress, and many pathophysiological mechanisms (inflammation, insulin resistance and metabolic adaptation) that develop during metabolic stress in peripartal period in dairy cows (Petrović i sar., 2018a). In this for cows during the difficult period around the partus there are numerous metabolic adaptations that arise as a result of gravity, partus and beginning of lactation (homeoretic processes), as well as the tendency of the organism to maintain homeostasis. All these processes inevitably lead to stress in the cows. The emergence of stress in cows is a consequence of the strengthening of homeotrophic, catabolic processes and the negative energy balance of cows at the beginning of lactation (Cincović 2013a).

The biggest problem with postpartum dairy cows is the imbalance between body reserves and milk production (Petrovic i sar., 2019a). The metabolic processes in the transition period are adapted to provide a sufficient amount of energy required as well as precursors for the synthesis of dairy compounds (Grummer 1995.; Overton and Valdrón, 2004). Consequently, the main feature of early lactation in dairy cows is the negative energy balance (NEBAL) and the state of metabolic stress that results from reduced intake of food, calving and starting lactation. Due to the negative energy balance, the organism increases its own reserves and enters the catabolism phase.

As a result, lipomobilization intensifies ketogenesis and lipogenesis in the liver, and as a consequence, the concentration of glucose, triglycerides and total blood cholesterol (Sevinc et al., 2003.; Đoković et al., 2007, 2009, 2010a).

In NEBAL conditions, the body consumes its own energy reserves, first of all glycogen reserves, then fat, and then protein. As a consequence, there is a fatty liver, a weight loss of various degrees, a decrease in production and reproductive abilities, and in some cases a death occurs (Đoković et al., 2014a).

Especially expressed is the increased mobilization of lipids from body depots in order to use fat for energy purposes, but cows become prone to the development of ketosis and fatty liver (Cincović et al., 2012; Đoković et al., 2014b). As a consequence of lipolysis in the fat tissue, the concentration of non-esterified fatty acids (NEFA) and betahydroxybutyrate (BHB) in the bloodstream increases. This phenomenon occurs as a result of endocrine and

metabolic changes, primarily due to the presence of insulin resistance (Cincović et al., 2014).

Stress resulting from a disbalance of energy metabolism with numerous endocrine, biochemical, haematological, immunological and other adaptations is called metabolic stress (Cincović 2013b). Metabolic stress in early lactation in dairy cows underlies many diseases (Cincovic et al., 2018).

The transition from the state of gravidity and drying in the period of early lactation (peripartal period) is a very strenuous process for the smooth functioning of the cow organism, and is the most critical for their health and productivity. Partus and lactation start metabolically burdening cows to the extremes, so numerous adaptations can be such that the number of cows with some of the diseases (fatty liver, ketosis, metritis, mastitis, left and **right** dislocation of **abomasum**, locomotor diseases) is increasing in this period (Cincovic 2013a; Petrović et al., 2019). There may also be a persistent decline in cow productivity through reduced milk production and poor maintenance of lactation, a significant loss of body condition after calving - BSC below 3.5 (3.25 to 3.75), the occurrence of reproductive disorders such as retained placenta and cyst ovary, causing great economic damage to farmers.

It should be noted that in all this good manufacturing practice on the farm affects the economic viability of production. Therefore, efficient reproduction on farms in dairy cows is of great economic importance. However, in modern farms, increased milk production along with poor farm management (poor nutrition or reproduction) can affect reduced fertility in animals. Namely, the selection in cattle breeding is very successful in the direction of increasing milk production. But the phenotype of milk yield is only 25%. The influence of paragenetic factors on milk properties, regardless of whether their nature is fixed (the breeding area of the years and the season of birth, season of calving, lactation in order) or continuous (age at first fertilization and calving) is very pronounced and significant, and it is necessary to include them in models for assessing the breeding value of dairy cows (Petrović et al., 2005, 2006, 2009, 2010, 2012; Bogdanović et al., 2012)

Unfortunately, with the increase in milk production, fertility decreases and, consequently, the number of animals that have partus, which is reflected in the profitability of production (Gábor et al., 2016).

The main goals of modern cattle breeding are the increase in the fertility of breeding throats, the prolongation of their exploitation time, the increase in profit and the cultivation of as many genetically high-quality offspring as possible. The achievement of these goals depends on the factors determining the genetic potential of the reproductive properties of the

reproductive throat and environmental factors (paragenetic factors), which enable the phenotypic expression of reproductive characteristics (Košarcic et al., 2003; Petrović et al., 2013a, b, c.).

Therefore, it is very important to understand the reproductive cycle physiology of dairy cows and its relationship with the metabolic status of cows in early lactation (Đoković et al., 2014a).

Cellular adaptation of dairy cows to peripartal metabolic stress (HSP70)

Milk production starts after calving of cows and is maintained artificially, by husband, for the next 305 days. In this period there are very significant changes in the metabolism and nutrition of cows, and the intensity and biological basis of these changes depend on the health of cows and milk production in the next lactation. The essence of metabolic changes occurring during the transition period (21 days before and 21 days after the partisans) is based on the fact that cows are exposed to negative energy balance during early lactation, that is, they can not enter through the food the amount of energy they need for maximum milk production. There are many reasons that lead to reduced food intake and a negative energy balance. Reduced food intake is the result of adaptation to the onset of lactation. On the other hand, in the cow there is a change in the main metabolic flows to maintain the lactation that follows. These processes lead to changes in the metabolism of cows, increasing the use of fat for energy purposes, in order to keep glucose for milk production. Also, in early lactation there is a deficit of vitamins and minerals, so their role in numerous metabolic processes is absent (Đoković et al., 2014c).

Namely, the basic changes in the metabolism of carbohydrates and fats in the period around calving and early lactation are: lower glycemic levels, increased gluconeogenesis, reduced glucose consumption in peripheral tissues, normal or decreased use of acetate, increased lipid mobilization from fat stores with elevated concentrations of non-esterified fatty acids (NEFA) and their increased use in peripheral tissues (Đoković 2010b)

These changes are followed by a series of endocrine changes such as insulin resistance, decreased insulin concentration (due to reduced food intake and decreased receptor sensitivity), decreased insulin-like growth factor IG and I IGF-I (due to a reduced anabolic effect of growth hormone in the peripheral tissue cow in spite of its elevated concentration), decreased thyroid hormone concentrations, catabolic effect of growth hormone on fatty tissue - increased growth hormone concentration (which allows increased use of nutrients in the mammary gland and leads to a decrease in insulin

sensitivity), elevated cortisol concentration (which helps lipomobilization and gluconeogenesis), increased glucagon levels.

During the transitional period in response to metabolic stress, numerous pathophysiological mechanisms (inflammation, insulin resistance and metabolic adaptation) are developed in dairy cows. Inflammation and insulin resistance are important pathophysiological mechanisms that develop during metabolic stress. Heat shock proteins have a significant influence in the regulation of both these processes in different animal species and humans. They help to clear the protein structure of the cell and its survival. However, if they find themselves extracellularly, they show proinflammatory effects and have to do with the development of insulin resistance and diabetes (Petrović et al., 2017).

In the organism of dairy cows numerous mechanisms are activated, the main role of which is to maintain all these processes within the physiological limits. In regulation of these processes, HSP 70 heat stress proteins play a key role.

Heat shock protein (HSP) are phylogenetically conserved and ubiquitous molecules, indicating their functional importance (Petrović et al., 2017).

Heat shock protein, HSP are chaperones necessary for the proper formation of the polypeptide chain and are responsible for its translocation in the cell. These proteins were found during exposure to heat stress, when their concentration and expression in the cells grew to explain their name (Cincović 2013a.; Petrović et al., 2016).

Heat shock proteins are synthesized in response to various forms of stress. Namely, their expression can be induced in several ways: physiological (growth factors and hormones), pathophysiological (infection, inflammation, ischemia, oxidative injuries and toxins), environmental conditions (heat stress and heavy metals) (Petrović i sar., 2018b).

They are traditionally classified according to their molecular weight. (Prohaszka and Fust, 2004.). According to the molecular mass we distinguish several types, so for example: 10 kDa (Hsp10), 20-30 kDa (Hsp27, HspB1), 40 kDa (Hsp40), 60 kDa (Hsp60), 70 kDa (Hsp70, Hsp71, Hsp72, Grp78, Hsx70), 90 kDa (Hsp90, Grp94) and 100 kDa (Hsp104, Hsp110). In cattle, four types of Hsp70 genes were identified, and IRNK for this protein was found in the tissue of different cell types and in the blood plasma (Agnew and Colditz 2008; Asea 2007).

Tavaria et al., (1996.) gave a first clarification of the nomenclature of the HSPA family and showed that the family of a human heat shock protein is composed of at least 12 members and many others agree with their allegations. Modification and extension was given by Kamping et el. (2009.),

where he provided updated guidelines for the nomenclature of the human HSPA family (HSP70), as well as HSPH (HSP110), HSPC (HSP90), DNAJ (HSP40) and HSPB (small HSP) and human chaperone families (HSP60 and CCT). Also, Kampinga et al., (2009.) stated that the guidelines for the nomenclature of human heat shock protein are also based on systemic gene symbols assigned by the HUGO Gene Nomenclature Committee (HGNC) and used as primary identifiers in databases such as Entrez Gene and Ensemble. The best known HSPs are: stress induced form HSP70 / HSP72 (HSPA1A), constitutive forms HSP70 / HSP73 / HSS73 (HSPA8), an endoplasmic reticulum form, Grp78 / BiP (HSPA5) and a form localized mainly in mitochondria HSP75 / mtHSP70 / mortalin / TRAP-1 (HSPA9) (Petrović et al., 2018c). In addition to them, and less familiar localization, there are: Hsp70-2 (HSPA1B); Hsp70-Hom / Hsp70t (HSPA1L); Hsp70-3 (HSPA2); Hsp70-6 / Hsp70B (HSPA6); HSP70-7 / Hsp70B (HSPA7), FLJ13874 / KIAA0417 (HSPA12A), RP23-32L15.1 / 2700081N06Rik (HSPA12B), Stch (HSPA13), HSP70-4 / HSP70L1 / MGC131990 (HSPA14) (Petrović et al., 2018a).

In the cells, the HSP70 family is the most induced HSP family in response to stress. HSP72 molecular weight of 72 kDa can represent up to 20% of the total cell protein and is very rapidly induced during cell stress after appropriate stimulation (Noble et al., 2008), especially in skeletal muscle cells (Madden et al., 2008).

Namely, the two most studied proteins in the HSP70 family are HSC73 and HSP72 (Beckmann et al., 1990). Sorger and Pelham (1987) have shown that HSC73, a heat shock protein molecule mass of 73 kDa, is synthesized in most cellular organisms and is only slightly inducible. Unlike HSC73, HSP72 is present in small amounts in ungraded cells, and is thought to be primarily stress-induced (Kiang and Tsokos 1998; Hartl 1996). During the action of various stress stimulus, the organism strives to meet increased demands during stress-related events in HSP72 synthesis (Black and Subject 1991).

HSP70 has the ability to exert completely opposite effects depending on its localization (Rodrigues-Krause et al., 2012). Namely, heat shock proteins have long been considered exclusively cytoplasmic proteins with certain functions that are limited to the intracellular part of the cell. However, an increasing number of studies have shown that they can be released into extracellular space (eHSP72) and have different effects on other cells (Titell 2005).

A high level of intracellular HSP72 synthesized in response to stress, occupies the cell and protects it through the role of molecular chaperon (Lindquist and Craig 1988). Said cytosolic inducible HSP70 can mediate

through cytoprotective, antiapoptotic and immunological regulatory effects, and is most studied.

The protective role of HSP70 is well documented, and it is interesting that HSP70-induced hyperthermia can provide protection against myocardial ischaemia, suggesting that HSP70 can be protected through cross-care (Cornelsson et al., 1994). Increased HSP70 expression in experimental models of stroke, sepsis, acute respiratory distress syndrome, renal insufficiency and myocardial ischemia is created to reduce bodily injury and in some cases improve survival (Jo et al., 2006; Weiss et al., 2002; Chen et al., 2003; Giffard and Yenari 2004). It has been shown that embryonic HSP70 plays a role in normal development (processes such as apoptosis, cell cycle regulation) and protects against stressors in sensitive embryonic stages (Luft and Dix 1999).

When it comes to extracellular HSP70, it plays a role of cytokine, an immunostimulatory role (helps synthesize proinflammatory cytokines) and improves antitumour control.

Extracellular eHSP70 comes from cells to the bloodstream from living cells exposed to stress through vesicular secretion, exosomes or lysosomes, and through intact lipid membranes that are independent of the transport of proteins through the endoplasmic reticulum-Golgi apparatus, but also passive pathways from necrotic cells and stress-stressed cells (Molvarec et al., 2007; J. Campisi et al., 2003). In a research by Campisi et al. (2003), extracellular heat shock proteins (eHSP), such as those belonging to the HSP family of 70 kDa (for example, HSP72) were presented to act as a "signal of danger" toward immune cells, promoting immune response and improving host defense.

The eHSP72 function is generally associated with the activation of the immune system (Whitham and Fortes 2008). For example, eHSP72 has been reported as an inductor of the microbicidal capacity of neutrophils (Ortega et al., 2006) and chemotaxis (Ortega et al., 2009), participates in the recruitment of NK killers (Horn et al., 2007) as well as in the production of cytokines in immune cells (Asea et al., 2000; Johnson and Fleshner 2006).

There is still no known HSP fraction in the bloodstream coming in one or the other way, and the role of extracellular HSP is contradictory. Namely, the concentration of HSP increases in various diseases, and due to the lack of HSP, metabolic syndrome occurs in humans (obesity, diabetes, cardiovascular disease and dyslipidemia) (Asea 2007; Chung et al., 2008; Krause and Rodrigues-Krause, 2011). Also, the increased concentration of extracellular HSP means better survival (Pittet et al., 2002).

Namely, although induction of iHSP72 reduces the production of cytokines, extracellular HSP (eHSP) can significantly increase the

production of proinflammatory cytokines (Breloer et al., 1999; Chen et al., 1999; Multhoff et al., 1999; Asea et al., 2000).

The concentration of HSP70 during gravidity and calving depends on numerous biological variables and physiology and pathology of calving (Molvarec et al., 2010).

Molvarec et al., (2007) found that the concentration of eHSP70 was lower in pregnancy than in non-pregnant women, which is consistent with our results (Petrović et al., 2016). Expression of Hsp72 mRNA in sheep myotome (intracellular) was elevated during lambing (Wu et al., 1996), as well as in amniotic fluid in women (extracellularly), which were conceived and produced on time (Chaiworapongsa et al., 2008).

Kristensen et al., (2004) have shown that there are numerous factors that influence the concentration of HSP70 in serum cows, such as age and stage of lactation. Although there was no statistically significant difference, plasma concentrations of HSP72 were higher in early lactation (the first 60 days) compared to the middle part of lactation. The concentration of HSP72 is significantly lower in the cows before the partus and in the first weeks after the partus, in order to grow. In dairy cows there is a positive correlation between extracellular and intracellular Hsp 72 values (Catalani et al., 2010). However, this indicates the existence of certain specificities in the regulation of extracellular Hsp72 in cows in the peripartal period. In cows in early lactation, a lower concentration of eHSP70 was found in the first two weeks after calving compared to 4 and 8 weeks (Petrović et al., 2016). These values, as well as their trend, agree with the results of Catalani et al. (2010) and Kristensen et al., (2004).

However, some studies have shown that caloric restriction, hypoglycaemia or hyperlipidemia (which occurs in early lactation) can regulate HSP expression in different parts of the body. Eitam et al., (2009) reported that an extended low-energy diet promoted cell-specific HSP response in cattle with a significant increase in HSP90, but unchanged levels of HSP70 mRNA in leukocytes and lower expression of HSP70 in somatic milk cells. Febbraio et al. (2004) showed that maintaining glucose availability during the exercise reduces the circulation response of HSP72 to healthy people. Creation of intracellular HSP72 under the effect of heat stress reduces insulin resistance and reduces fat accumulation in hepatocytes (Morino et al., 2008). HSP72 concentrations in leukocytes and plasma increased rapidly after calving and correlated with NEFA, glucose, and TNF α (Catalani et al., 2010).

In a small number of studies, the association of peripartal metabolic stress with the values of chaperones was examined. The NEFA concentration in peripartal period shows a positive correlation with the NSP72

concentration (Catalani et al., 2010). Cincović and Belić (2014) showed that the concentration of NSP70 was significantly higher in weeks after calving compared to a week before calving. A higher concentration of NEFA and BHB (beta-hydroxybutyrate) was found in the first and second weeks after calving compared to other periods. The concentration of NSP70 positively correlates with NEFA and BHB values. Partial correlation shows that ties are stronger in the first and second weeks after calving, which is the period when lipid mobilization and ketogenesis are most pronounced. The concentration of NSP70 in the first two weeks after calving is dependent on the level of lipid mobilization and ketogenesis. Metabolic stress, characterized by lipid mobilization and ketogenesis, increases the blood NSP70 concentration during early lactation.

CONCLUSIONS

Hsp70 shows significant relationships with the pathophysiological mechanisms dominant in cows in early lactation, such as inflammatory response and insulin resistance.

Consequently, there is a presumption that the indicators of metabolic stress can affect the concentration of Hsp70 in serum of cows.

In the future, Hsp70 can be a significant indicator that can be used to evaluate the metabolic adaptation of cows in the peripartal period.

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REFERENCES

- Miloš Ž. PETROVIĆ, Radojica D. ĐOKOVIĆ, Marko R. CINCOVIĆ, Branislava BELIĆ, Milun D. PETROVIĆ, Vladimir KURĆUBIĆ, Zoran Ž. ILIĆ, Miodrag Radinović (2018a): Family of heat shock proteins of 70 kDa in the peripartal period in dairy cows. IX International Scientific Agriculture Symposium "Agrosym 2018", Jahorina, October 04 - 07, 2018., p. 1063. ISBN 978-99976-718-5-1.
- Cincović M. R. (2013a): Patofiziološka procena peripartalnog metaboličkog stresa kod visokoproduktivnih krava upotrebom endokrinih i

- metaboličkih kriterijuma. Doktorska disertacija. Univerzitet u Novom Sadu, Poljoprivredni fakultet, Departman za veterinarsku medicinu.
- Miloš Ž. Petrović, Radojica Đoković, Marko Cincović, Branislava Belić, Miodrag Radinović, Milun D. Petrović, Vladimir Kurćubić, Zoran Ž. Ilić (2019): Uticaj metaboličkog statusa na reproduktivnu efikasnost mlečnih krava u peripartalnom period. XXIV Savetovanje o biotehnologiji sa međunarodnim učešćem, Zbornik radova, str. 639-646, ISBN 978-86-87611-68-9; ISBN 978-86-87611-69-6 (niz); COBISS.SR-ID 274576652; 15-16. mart 2019. godine, Čačak.
- GRUMMER R.R. (1995): Impact of changes in organic nutrient metabolism on feeding the transition dairy cows. *J. Anim. Sci.*, 73: 2820-2833.
- OVERTON T.R., WALDRON. M.R. (2004): Nutritional management of transition dairy cows: Strategies to optimize metabolic health. *J. Dairy Sci.*, 87: E105-E119.
- SEVINC M., BASOGLU A., GUZELBEKTA H. (2003): Lipid and lipoprotein levels in dairy cows with fatty liver. *Turk. J. Vet. Anim. Sci.*, 27: 295-299.
- ĐOKOVIĆ R., ŠAMANC H., JOVANOVIĆ M., NIKOLIC Z. (2007): Blood concentrations of thyroid hormones and lipids in the liver in dairy cows in transitional period. *Acta Vet. Brno*, 76:525-532.
- ĐOKOVIĆ R., ILIĆ Z., BOGOSAVLJEVIĆ-BOŠKOVIĆ S., PETROVIĆ M. (2009): The functional state of liver cells in dairy cows in postpartal period and during lactation. *Contemporary agriculture (Novi Sad)*, 58 (3-4)37-43.
- ĐOKOVIĆ R., ILIĆ Z., KURĆUBIĆ V., DOSKOVIĆ V. (2010a): The values of organic and inorganic blood parameters in dairy cows during the peripartal period. *Contemporary agriculture (Novi Sad)*, 59 (1-2)30-36.
- Радочица Д. Ђоковић, Нектариос Д. Гиединис, Стаматис Аргироудис, Јован А. Бојковски, (2014а): Здравствена заштита преживара. 24-26. Чачак, Србија, Агрономски факултет у Чачку.
- Cincović M.R., Belić B., Radojičić B., Hristov S., Đoković R. (2012): Influence of lipolysis and ketogenesis to metabolic and hematological parameters in dairy cows during periparturient period. *Acta veterinaria (Beograd)*, 62(4), 429-444.
- Djoković R. Cincović, M., Kurcubic, V., Petrović, M., Lalović, M., Jašović, B., Stanimirović, Z. (2014b). Endocrine and Metabolic Status of Dairy

- Cows during Transition Period. Thai Journal of Veterinary Medicine, 44(1), 59-66.
- Cincović M.R., Belić B., Đoković R., Toholj B., Hristovska T., Delić B., Došenović M. (2014): Insulin resistance in cows during dry period and early lactation. Contemporary agriculture, 63, 1-2, 98-105.
- Cincović M.R. (2013b): Upotreba indikatora insulinske rezistencije u proceni metaboličkog statusa krava u ranoj laktaciji. Specijalistički rad. Univerzitet u Beogradu, Fakultet veterinarske medicine, Beograd.
- Марко Р. Цинцовић, Радојица Ђоковић, Бранислава Белић, Милош Петровић, Ивана Лакић (2018): Кетоза и инсулинска резистенција код крава. Зборник радова и кратких садржаја. 29. Саветовање ветеринара Србије, стр 238-242, 13-16 септембар, Златибор ISBN 978-86-83115-35-8, COBBIS.SR-ID 267576844.
- PETROVIĆ D.M., SKALICKI Z., BOGDANOVIĆ V., PETROVIĆ M.M., KURĆUBIĆ V. (2005): The Effect of Paragenetic Factors on Performance Traits in Complete Lactations in Simmental Cows. 8th International Symposium Modern Trends In Livestock Production. Belgrade Zemun, Serbia and Montenegro, 5.-8.10.2005. Biotechnology in Animal Husbandry 21 (5-6), p 7-12.
- PETROVIĆ D.M., ĐOKOVIĆ R., BOGOSAVLJEVIĆ-BOŠKOVIĆ SNEŽANA., KURĆUBIĆ V. (2006): Uticaj paragenetskih faktora na proizvodne osobine standardnih laktacija kod krava simentalske rase. Savremena poljoprivreda, Vol.55, 1-2, Str. 138-143, Novi Sad.
- PETROVIĆ D.M., SKALICKI Z., PETROVIĆ M.M., BOGDANOVIĆ V. (2009): The Effect of Systematic Factors on Milk Yield in Simmental Cows Over Complete Lactations. Biotechnology in Animal Husbandry 25(1-2), p 61-71, Belgrade-Zemun.
- PETROVIĆ D.M., BOGDANOVIĆ V., PETROVIĆ M.M., SNEŽANA BOGOSAVLJEVIĆ-BOŠKOVIĆ (2010): Uticaj paragenetskih faktora na proizvodnju 4% mast-korigovanog mleka u celim i standardnim laktacijama. XV savetovanje o biotehnologiji. Zbornik radova Vol. 15(17), str. 585-590. Agronomski fakultet, Čačak.
- M.D. PETROVIĆ, Z. SKALICKI, V. BOGDANOVIĆ, M.M. PETROVIĆ, S. BOGOSAVLJEVIĆ-BOŠKOVIĆ, R. ĐOKOVIĆ, S. RAKONJAC (2012): The Effect of Geographical Region on Lifetime Milk Yield in Simmental

Cows. Proceedings of the First International Symposium on Animal Science. November 8-10th, Belgrade, Serbia. Book I, p. 103-110. ISBN 978-86-7834-164-9, ISBN 978-86-7834-166-3.

BOGDANOVIĆ V., ĐEDOVIĆ R., STANOJEVIĆ D., PETROVIĆ D.M., BESKOROVAJNI R., RUŽIĆ-MUSLIĆ D., PANTELIĆ V. (2012): Regional Differences in Expression of Milk Production Traits in Simmental Cows. Proceedings of the First International Symposium on Animal Science. November 8-10th, Belgrade, Serbia. Book I, p. 223-230.

Gábor, G., Balogh, O.G., Kern, L., Gábor, P.R. and Fébel, H. (2016): Nutrition, Metabolic Status and Reproductive Efficiency in Dairy Herds. Open Journal of Animal Sciences, 5, 75-84.

Košarčić D, Košarčić S, Grubac S. (2003): Uloga i značaj biotehnologija u reprodukciji domaćih životinja. "Savremena poljoprivreda" vol. 52, 3-4, str. 209-213, Novi Sad

M.D. Petrović, M.M. Petrović, V. Bogdanović, S. Bogosavljević-Bošković, R. Đedović, S. Rakonjac (2013a): Effect Of Fixed And Continuous Non-Genetic Factors On Length Of Service Period In Simmental Cows. Proceedings of the 10th International Symposium Modern Trends in Livestock Production. Belgrade, Serbia, October 2-4, 2013. p. 48-56. ISBN 978-86-82431-69-5.

Petrović M.D., Bogdanović V., Bogosavljević-Bošković S., Petrović M.M., Đoković R., Rakonjac S., Dosković V. (2013b): Effect of Fixed and Continuous Non-genetic Factors on Calf Birth Weight. 23rd International symposium "New Technologies in Contemporary Animal Production", Novi Sad (Serbia), 19-21 Jun, 2013, p. 50-52, ISBN 978-86-7520-271-4.

M. D. Petrović, M.M. Petrović, V. Bogdanović, R. Đedović, R. Đoković, V. Dosković, S. Rakonjac (2013c): Effect of Fixed and Continuous Non-genetic Factors on length of Calving Interval in Simmental cows. Journal of Mountain Agriculture on the Balkans, Vol 16, 4. p. 880-895. Published by Research Institute of Mountain Stockbreeding and Agriculture Trojan, Bulgaria, pp. ISSN 1311-0489.

Радојица Д. Ђоковић, Марко Р. Џинцовић, Бранислава М. Белић (2014c): Физиологија и патофизиологија метаболизма крва у

- перипарталном периоду. Помоћни Уџбеник, Универзитет у Новом Саду, Департман за ветеринарску медицину.
- Đoković R.D. (2010b.): Endokrini status mlečnih krava u peripartalnom periodu. Agronomski fakultet, Čačak.
- Милош Петровић, Марко Р. Цинковић, Радојица Ђоковић, ЈожеСтарич, Бранислава Белић, Јожица Јежек (2017): Повезаност протеина тоplotног шока HSP70 са инфламацијом и инсулинском резистенцијом - импликације код млечних крава. Зборник радова и кратких садржаја. 28. Саветовање ветеринара Србије, стр 153-157, 7-10 септембар, Златибор ISBN 978-86-83115-32-7, COBBIS.SR-ID 243984140.
- Miloš Petrović, Marko R. Cincović, Branislava Belić, Radojica Đoković, Jože Starič, Jožica Ježek (2016): Koncentracija protein toplotnog stresa (Heat shock protein 70, Hsp70) u krvnom serumu kod krava u ranoj laktaciji. 27. Savetovanje veterinarara Srbije, Zlatibor 8 - 11. 09. 2016., str 235-238. ISBN 978-86-83115-30-3
- Miloš Ž. PETROVIĆ, Radojica D. ĐOKOVIĆ, Marko R. CINCOVIĆ, Branislava BELIĆ, Milun D. PETROVIĆ, Vladimir KURČUBIĆ, Zoran Ž. ILIĆ, Miodrag Radinović (2018b). The function of heat shock protein hsp70 in dairy cows in early lactation. IX International Scientific Agriculture Symposium "Agrosym 2018", Jahorina, October 04 - 07, 2018., p. 1064. ISBN 978-99976-718-5-1. COBISS.RS-ID 7679512
- Prohaszka Z, Fust G (2004): Immunological aspects of heat-shock proteins-the optimum stress of life. *Mol Immunol* 41:29–44
- Agnew L.L., Colditz I.G. (2008): Development of a method of measuring cellular stress in cattle and sheep. *Veterinary Immunology and Immunopathology*, 123: 197–204.
- Asea A. (2007): Mechanisms of HSP72 release. *J. Biosci.*, 32(3): 579–584.
- Tavaria M, Gabriele T, Kola I, Anderson RL (1996): A hitchhiker's guide to the human Hsp70 family. *Cell Stress Chaperones* 1:23–28.
- Kampinga HH, Hageman J, Vos MJ et al (2009): Guidelines for the nomenclature of the human heat shock proteins. *Cell Stress Chaperones* 14:105–111.

- Miloš Petrović, Radojica Đoković, Milun D. Petrović, Vladimir Kurćubić, Marko Cincović, Branislava Belić, Zoran Ž. Ilić, Neđeljko Karabasil (2018c). Importance of intracellular and extracellular protein hsp70 in peripartal period in dairy cows. 7th International Symposium on Agricultural Sciences "AgroReS 2018" February 28 – March 2, 2018; Banja Luka, Bosnia and Herzegovina. Str 109. CIP - 631(048.3)(0.034.2). ISBN 978-99938-93-45-5. COBISS.RS-ID 7238680
- Noble EG, Milne KJ, Melling CW (2008): Heat shock proteins and exercise: a primer. *Appl Physiol Nutr Metab* 33(5):1050 – 1065. doi:10.1139/H08-069
- Madden LA, Sandstrom ME, Lovell RJ, McNaughton L (2008): Inducible heat shock protein 70 and its role in preconditioning and exercise. *Amino Acids* 34(4):511 – 516. doi:10.1007/s00726-007-0004-7
- Beckmann R. P, L. A. Mizzen and W. J. Welch (1990): Interaction of hsp70 with newly synthesized proteins: implications for protein folding and assembly events. *Science* 248, 850-854
- Sorger P. K and H. R. B. Pelham (1987): Cloning and expression of a gene encoding HSC73, the major HSP70- like protein in unstressed rat cells. *EMBO J* 6, 993-998
- Kiang JG, Tsokos GC. (1998): Heat shock protein 70 kDa: molecular biology, biochemistry, and physiology. *Pharmacol Ther* 80: 183–201.
- Hartl FU. (1996): Molecular chaperones in cellular protein folding. *Nature* 381: 571–579.
- Black A. R and J. R. Subject (1991): Systemic effects of stress. The biology and physiology of the heat shock and glucose-regulated stress protein systems. *Methods Arch Exp Pathol* 15, 126-166.
- J. Rodrigues-Krause, M. Krause, C. O'Hagan et al., (2012): "Divergence of intracellular and extracellular HSP72 in type 2 diabetes: does fat matter?" *Cell Stress and Chaperones*, vol. 17, no. 3, pp. 293– 302.
- Tytell M (2005): Release of heat shock proteins (Hsps) and the effects of extracellular Hsps on neural cells and tissues. *Int J Hyperthermia* 21(5):445 – 455. doi:10.1080/02656730500041921
- Lindquist S, E. A. Craig (1988): The heat-shock proteins. *Annu Rev Genet* 22, 631-677.

- Cornelusson R, W. Spiering, J. H. G. Webers, L. G. DeBruin, R. S. Reneman, G. J. Van der Vusse and L. H. E. H. Snoeckx (1994): Heat shock improves ischemic tolerance of hypertrophied rat hearts. *Am J Physiol* 267, H1941-H1947.
- Jo SK, Ko GJ, Boo CS, Cho WY, Kim HK (2006): Heat preconditioning attenuates renal injury in ischemic ARF in rats: role of heat-shock protein 70 on NF-kappaB-mediated inflammation and on tubular cell injury. *J Am Soc Nephrol* 17:3082–3092.
- Weiss YG, Maloyan A, Tazelaar J, Raj N, Deutschman CS (2002): Adenoviral transfer of HSP-70 into pulmonary epithelium ameliorates experimental acute respiratory distress syndrome. *J Clin Invest* 110:801–806.
- Chen HW, Hsu C, Lu TS, Wang SJ, Yang RC (2003): Heat shock pretreatment prevents cardiac mitochondrial dysfunction during sepsis. *Shock* 20:274–279.
- Giffard RG, Yenari MA (2004): Many mechanisms for hsp70 protection from cerebral ischemia. *J Neurosurg Anesthesiol* 16:53–61.
- Luft JC, Dix DJ (1999): Hsp70 expression and function during embryogenesis. *Cell Stress Chaperones* 4:162–170.
- Molvarec A, Rigó J Jr, Nagy B, Walentin S, Szalay J, Füst G, Karádi I, Prohászka Z (2007): Serum heat shock protein 70 levels are decreased in normal human pregnancy. *J Reprod Immunol* 74:163–169.
- J. Campisi, T. H. Leem, and M. Fleshner (2003): “Stress-induced extracellular Hsp72 is a functionally significant danger signal to the immune system,” *Cell Stress & Chaperones*, vol. 8, no. 3, pp. 272–286.
- Whitham M, Fortes MB (2008): Heat shock protein 72: release and biological significance during exercise. *Front Biosci* 13:1328 – 1339.
- Ortega E, Giraldo E, Hinchado MD, Martinez M, Ibanez S, Cidoncha A, Collazos ME, Garcia JJ (2006): Role of Hsp72 and norepinephrine in the moderate exercise-induced stimulation of neutrophils' microbicide capacity. *Eur J Appl Physiol* 98(3):250 – 255. doi:10.1007/s00421-006-0269-7.
- Ortega E, Hinchado MD, Martin-Cordero L, Asea A (2009): The effect of stress-inducible extracellular Hsp72 on human neutrophil chemotaxis:

- a role during acute intense exercise. *Stress* 12(3):240 –249. doi:10.1080/10253890802309853.
- Horn P, Kalz A, Lim CL, Pyne D, Saunders P, Mackinnon L, Peake J, Suzuki K (2007): Exercise-recruited NK cells display exercise-associated eHSP-70. *Exerc Immunol Rev* 13:100 – 111.
- Asea A, Kraeft SK, Kurt-Jones EA, Stevenson MA, Chen LB, Finberg RW, Koo GC, Calderwood SK (2000): HSP70 stimulates cytokine production through a CD14-dependant pathway, demonstrating its dual role as a chaperone and cytokine. *Nat Med* 6(4):435 – 442. doi:10.1038/74697.
- Johnson JD, Fleshner M (2006): Releasing signals, secretory pathways, and immune function of endogenous extracellular heat shock protein 72. *J Leukoc Biol* 79(3):425 – 434. doi:10.1189/jlb.0905523.
- Asea A, (2007): Mechanisms of HSP72 release. *J. Biosci.*, 32 (3) 579-584.
- Jason Chung, Anh-Khoi Nguyen, Darren C. Henstridge, Anna G. Holmes, M. H. Stanley Chan, Jose L. Mesa, Graeme I. Lancaster, Robert J. Southgate, Clinton R. Bruce, Stephen J. Duffy, Ibolya Horvath, Ruben Mestril, Matthew J. Watt, Philip L. Hooper, Bronwyn A. Kingwell, Laszlo Vigh , Andrea Hevener, and Mark A. Febbraio (2008): HSP72 protects against obesity-induced insulin resistance. *Proc Natl Acad Sci U S A*. 2008; 105(5):1739-44. doi: 10.1073/pnas.0705799105
- Krause M, Rodrigues-Krause Jda C (2011): Extracellular heat shock proteins (eHSP70) in exercise: possible targets outside the immune system and their role for neurodegenerative disorders treatment. *Med Hypotheses* 76(2):286 – 290. doi:10.1016/j.mehy.2010.10.025
- Jean-François Pittet; H. Lee; Diane Morabito; M. B. Howard; William J. Welch; Robert C. Mackersie (2002): Serum Levels of Hsp 72 Measured Early after Trauma Correlate with Survival. *The Journal of Trauma: Injury, Infection, and Critical Care*. 52(4):611-617).
- Breloer M, Fleischer B, Von Bonin A. (1999): In vivo and in vitro activation of T cells after administration of Ag-negative heat shock proteins. *J Immunol* 162: 3141–3147.
- Chen W, Syldath U, Bellmann K, Burkart V, Kolb H. (1999): Human 60-kDa heat-shock protein: a danger signal to the innate immune system. *J Immunol* 162: 3212–3219.

- Multhoff G, Mizzen L, Winchester CC, et al. (1999): Heat shock protein 70 (Hsp70) stimulates proliferation and cytolytic activity of natural killer cells. *Exp Hematol* 27: 1627–1636.
- Attila Molvarec & Lilla Tamási & György Losonczy & Krisztina Madách & Zoltán Prohászka & János Rigó Jr. (2010): Circulating heat shock protein 70 (HSPA1A) in normal and pathological pregnancies. *Cell Stress and Chaperones* (2010) 15:237–247.
- Molvarec A, Rigó J Jr, Nagy B, Walentin S, Szalay J, Füst G, Karádi I, Prohászka Z (2007): Serum heat shock protein 70 levels are decreased in normal human pregnancy. *J Reprod Immunol* 74:163–169.
- Wu WX, Derks JB, Zhang Q, Nathanielsz PW (1996): Changes in heat shock protein-90 and -70 messenger ribonucleic acid in uterine tissues of the ewe in relation to parturition and regulation by estradiol and progesterone. *Endocrinology* 137:5685–5693.
- Chaiworapongsa T, Erez O, Kusanovic JP, Vaisbuch E, Mazaki-Tovi S, Gotsch F, Than NG, Mittal P, Kim YM, Camacho N, Edwin S, Gomez R, Hassan SS, Romero R (2008): Amniotic fluid heat shock protein 70 concentration in histologic chorioamnionitis, term and preterm parturition. *J Matern Fetal Neonatal Med* 21:449–461.
- Kristensen TN, Løvendahl P, Berg P, Loeschcke V (2004): Hsp72 is present in plasma from Holstein Friesian dairy cattle, and the concentration level is repeatable across days and age classes. *Cell Stress Chaperones* 9:143–149.
- Catalani E., Amadori M., Vitali A., Bernabucci U., Nardone A., Lacetera N. (2010): The Hsp72 response in peri-parturient dairy cows: relationships with metabolic and immunological parameters. *Cell Stress & Chaperones*, 15(6): 781-790.
- Eitam H, Brosh A, Orlov A, Izhaki SA (2009): Caloric stress alters fat characteristics and HSP70 expression in somatic cells of lactating beef cows. *Cell Stress Chaperones* 14:173–82.
- Febbraio MA, Mesa JL, Chung J, Steensberg A, Keller C, Nielsen HB, Krstrup P, Ott P, Secher NH, Pedersen BK (2004): Glucose ingestion attenuates the exercise-induced increase in circulating heat shock protein 72 and heat shock protein 60 in humans. *Cell Stress Chaperones* 9:390–396.

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- Morino S, Kondo T, Sasaki K, Adachi H, Suico MA, Sekimoto E, Matsuda T, Shuto T, Araki E, Kai H (2008) Mild electrical stimulation with heat shock ameliorates insulin resistance via enhanced insulin signalling. PLoS ONE 3:e4068.
- Cincovic M, Belic B. (2014): Concentration of blood Hsp70 and its relation With lipid mobilisation and ketogenesis in dairy cows during periparturient period, Contemporary agriculture. 63, 1-2, 92-7.

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